

# Computational Modeling of the Cardiovascular System

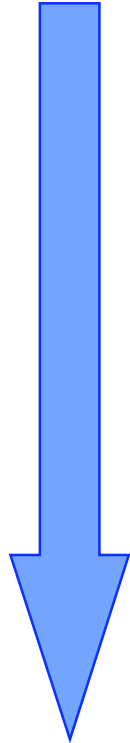
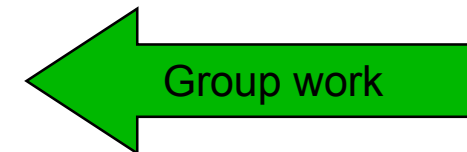
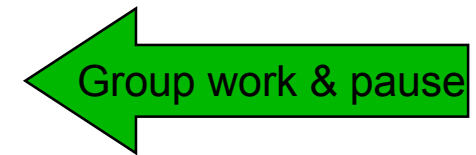
## Electrophysiological Modeling of Membranes and Ion Channels



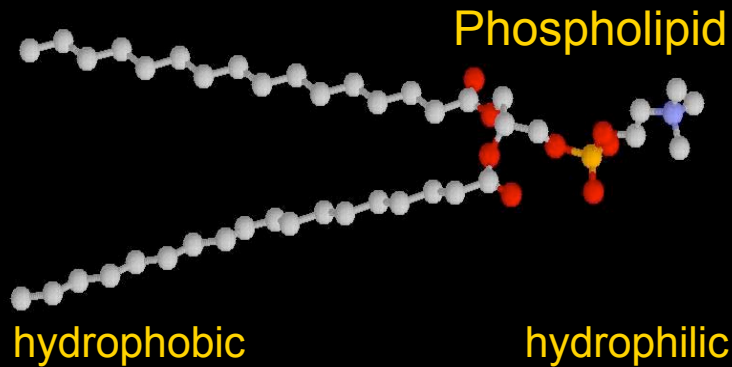
**Frank B. Sachse**, University of Utah

# Overview

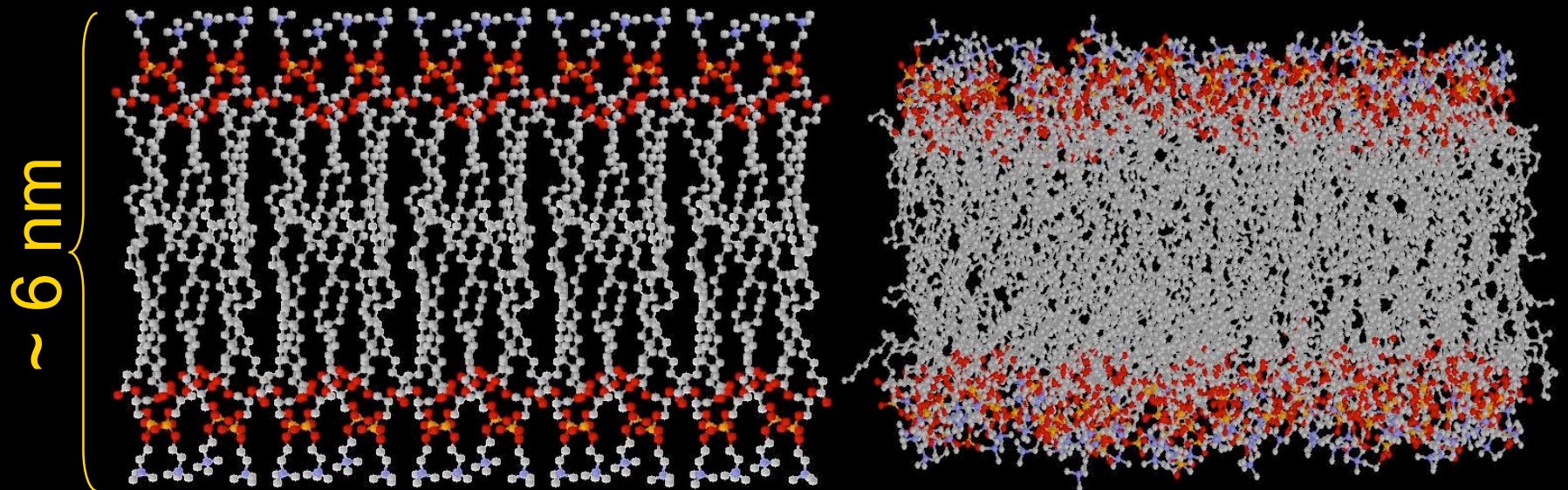
- Membranes
  - Resistor-Capacitor Model
  - Equations of Nernst and Goldman-Hodgkin-Katz
  - Squid Axon Model
- Channel Modeling
  - Structure
  - Experimental Studies
  - Markov Modeling
- Numerical Methods



# Molecular Structure of Phospholipid Bilayers



- Nitrogen
  - Oxygen
  - Phosphor
  - Carbon
- (Hydrogen not represented)



(Structure data from Heller et al, J. Phys. Chem., 1993)

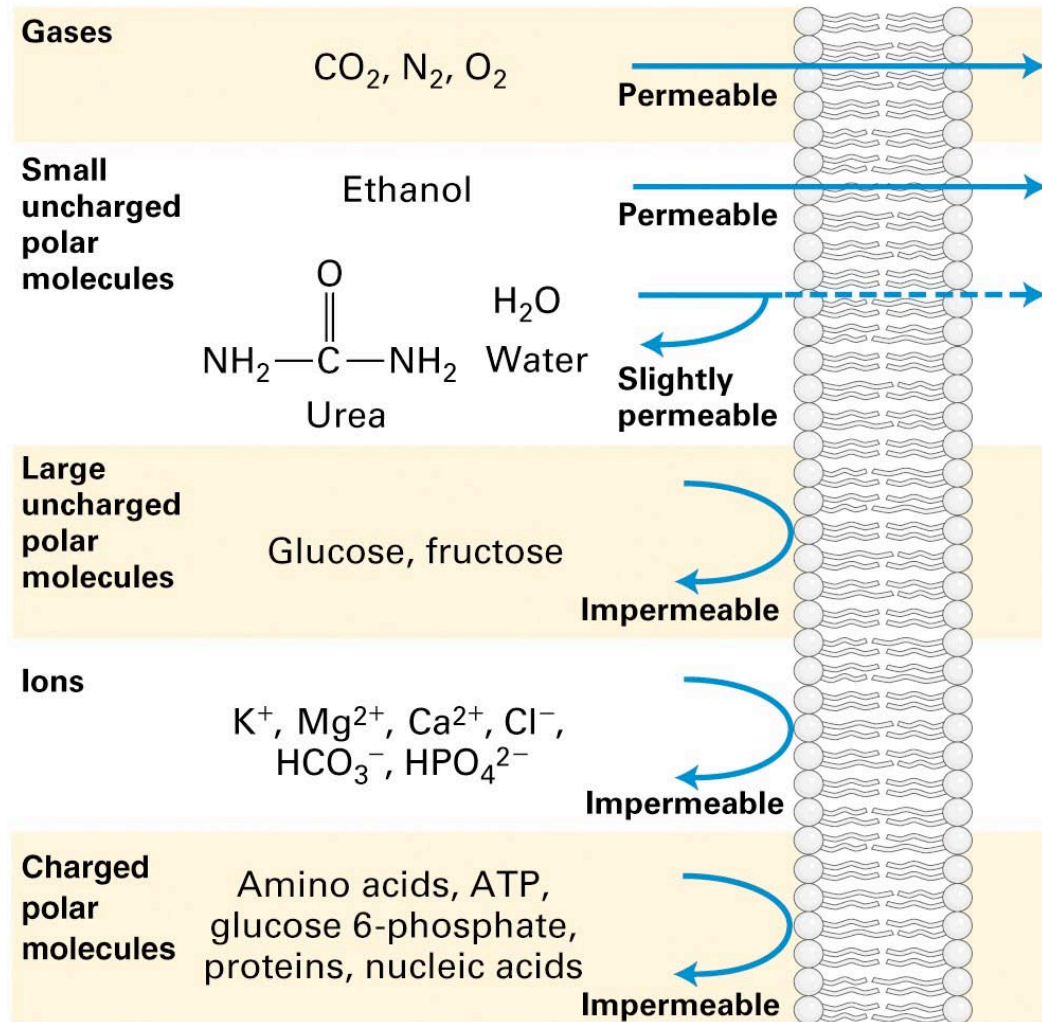
# Phospholipid Bilayers

- Plasma membrane
- Membrane of organelle

Selective permeability

Transmembrane proteins responsible for transport:

- Ion Channels
- Pumps
- Exchangers



(Lodish et al., Molecular Cell Biology, Fig. 7-1, 2004)

## Modeling of Membrane: Resistor-Capacitor Circuit

$$C_m = \frac{Q}{V_m}$$

$C_m$  : membrane capacity [F]

$Q$  : electrical charge [As]

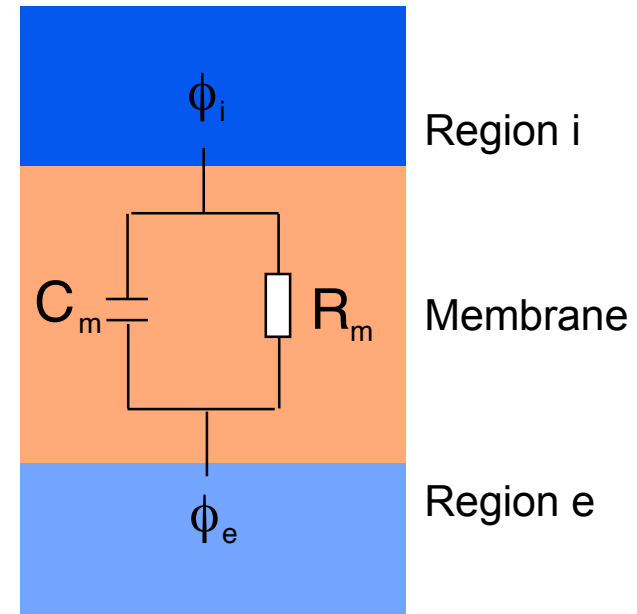
$V_m = \phi_i - \phi_e$  : voltage over membrane [V]

$$\frac{d}{dt} V_m = \frac{d}{dt} \frac{Q}{C_m} = \frac{I_m}{C_m}$$

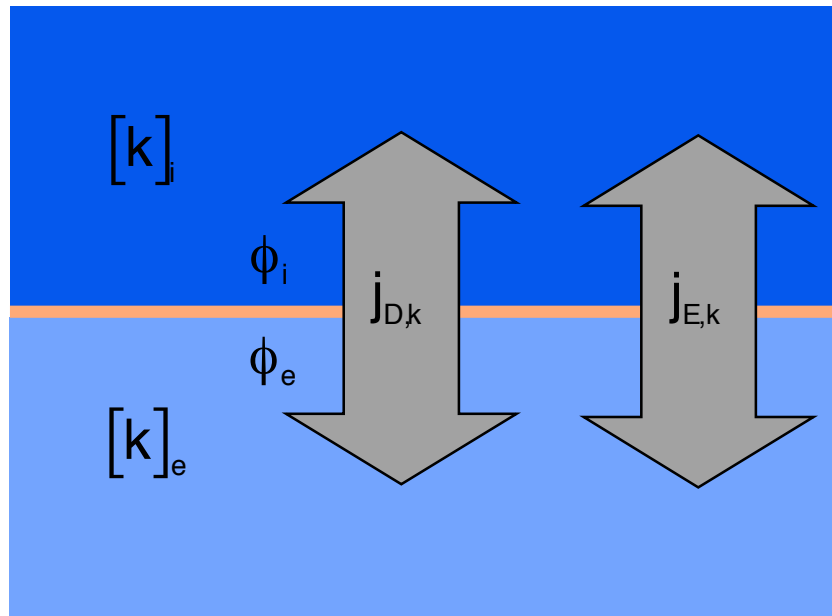
$I_m$  : Current through membrane [A]

$$R_m = -\frac{V_m}{I_m}$$

$R_m$  : Resistance of membrane [ $\Omega$ ]



# Modeling of Membrane: Nernst Equation



Region i

Membrane

- permeable for ion type k
- homogeneous, planar, infinite

Region e

$[k]_i$  : Concentration of k in region i

$\phi_i$  : Potential in region i

$j_{D,k}$  : Ionic current by diffusion

$[k]_e$  : Concentration of k in region e

$\phi_e$  : Potential in region e

$j_{E,k}$  : Ionic current by electrical forces

# Modeling of Membrane: Nernst Potential

In Equilibrium

$$j_{E,k} + j_{D,k} = 0$$

Malmivuo, Plonsey  
3.2.3

$$V_{m,k} = \phi_i - \phi_e = -\frac{RT}{z_k F} \ln \frac{[k]_i}{[k]_e}$$

$k$  : Ion type

$V_{m,k}$  : Nernst potential [V]

$R$  : Gas constant [J/mol/K]

$T$  : Absolute temperature [K]

$z_k$  : Valence

$F$  : Faraday's constant [C/mol]

$[k]_i$  : intracellular concentration of ion type  $k$  [M]

$[k]_e$  : extracellular concentration of ion type  $k$  [M]

## Modeling of Membrane: Nernst Equation - Example

Nernst equation explains measured transmembrane voltage of animal and plant cells

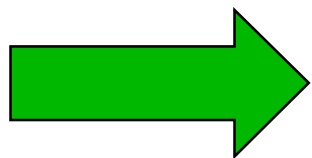
For potassium (monovalent cation) at temperatures of 37°C:

$$V_{m,K} = -\frac{310K}{+1} \frac{R}{F} \ln \frac{[K]_i}{[K]_o} = -61mV \log \frac{[K]_i}{[K]_e}$$

For typical intra- and extracellular concentrations:

$$[K]_i = 150 \text{ mM}$$

$$[K]_e = 5.5 \text{ mM}$$

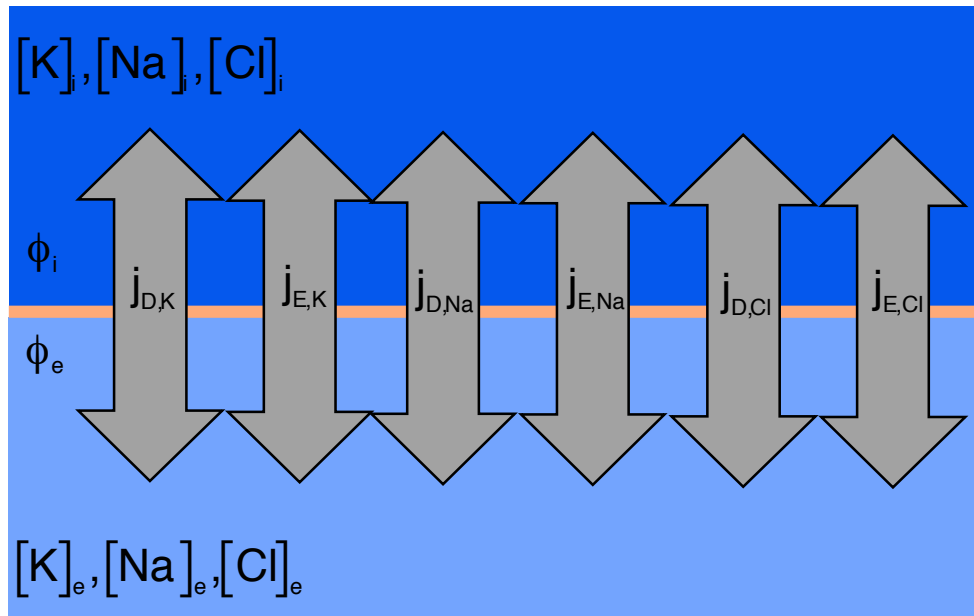


$$V_{m,K} = -88mV$$

Commonly, several types of ions are contributing to transmembrane voltage!



# Modeling of Membrane: Goldman-Hodgkin-Katz Equation



Region i

Membrane

- permeable for Na, K, Cl ions
- homogeneous, planar, infinite

Region e

$[Na]_i, [K]_i, [Cl]_i$  : Concentration in region i

$\phi_i$  : Potential in region i

$j_{D,Na}, j_{D,K}, j_{D,Cl}$  : Ionic current by diffusion

$[Na]_e, [K]_e, [Cl]_e$  : Concentration in region e

$\phi_e$  : Potential in region e

$j_{E,Na}, j_{E,K}, j_{E,Cl}$  : Ionic current by electrical forces



## Modeling of Membrane: Goldman-Hodgkin-Katz Equation

$$V_m = \phi_i - \phi_e = -\frac{RT}{F} \ln \frac{P_K [K]_i + P_{Na} [Na]_i + P_{Cl} [Cl]_e}{P_K [K]_e + P_{Na} [Na]_e + P_{Cl} [Cl]_i}$$

$V_m$  : Equilibrium voltage over membrane [V]

$R$  : Gas constant [J/mol/K]

$T$  : Absolute Temperature [K]

$F$  : Faraday constant [C/mol]

$[K]_i, [Na]_i, [Cl]_i$  : Intracellular concentrations [M]

$[K]_e, [Na]_e, [Cl]_e$  : Extracellular concentrations [M]

$P_K, P_{Na}, P_{Cl}$  : Permeabilites [cm/s]

# Hodgkin and Huxley: Measurements

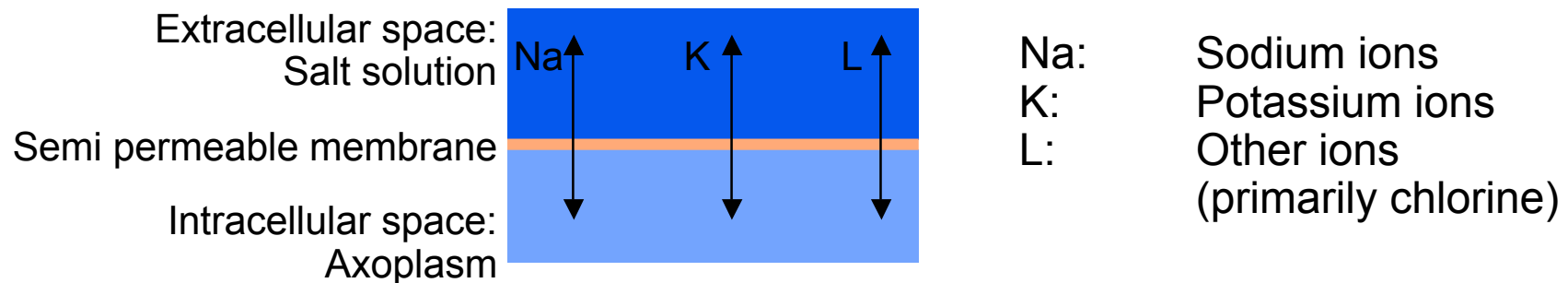
Measurement and mathematical modeling of electrophysiological properties of cell membrane (published 1952, Nobel prize 1963)

“Giant” axon from squid with ~0.5 mm diameter

Techniques

- Space clamp
- Voltage clamp

Simplifications:



# Hodgkin-Huxley: Clamp Techniques

## • Space Clamp

Electrophysiological properties are independent of  $x$

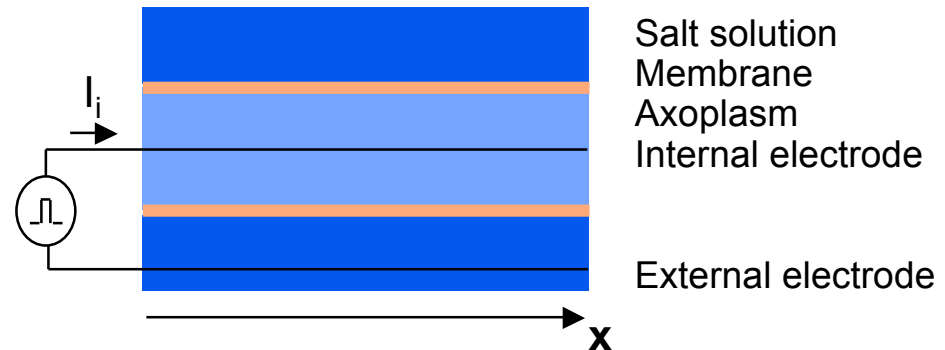
$$I_m = I_i + C_m \frac{d}{dt} V_m$$

$I_i$  : Injected current [A]

$I_m$  : Current through membrane [A]

$C_m$  : Membrane capacitor [F]

$V_m$  : Membrane voltage [V]



## • Voltage Clamp

Voltage  $V_m$  is kept constant by injection of current  $I_i$



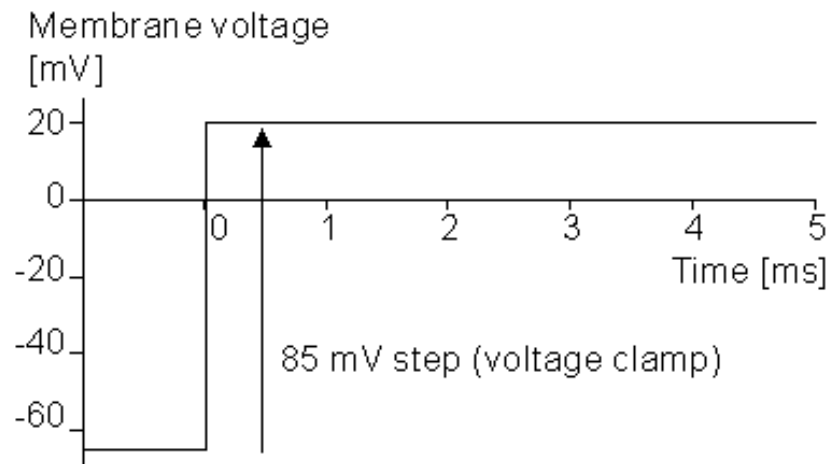
neglect of capacitive currents



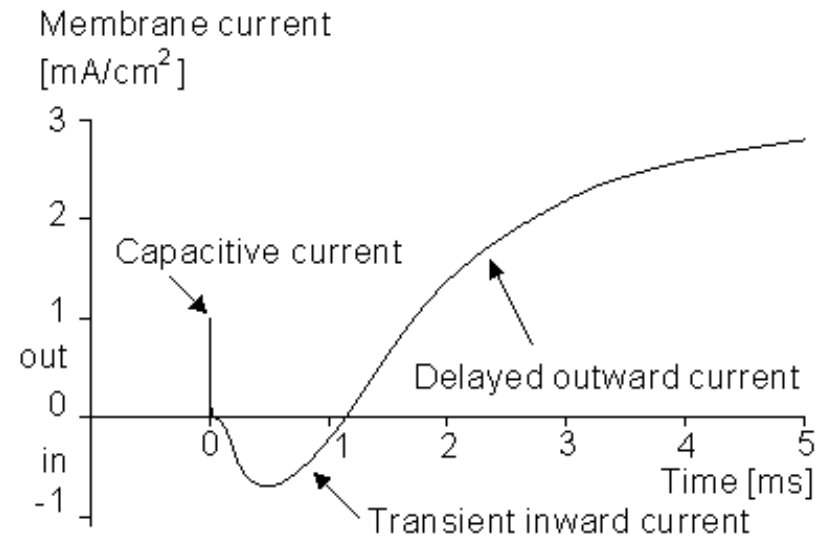
$$I_m = I_i$$



# Hodgkin-Huxley: Voltage clamping

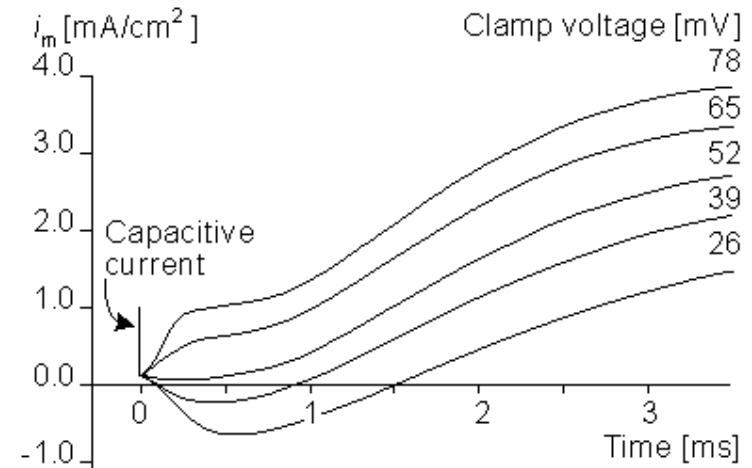
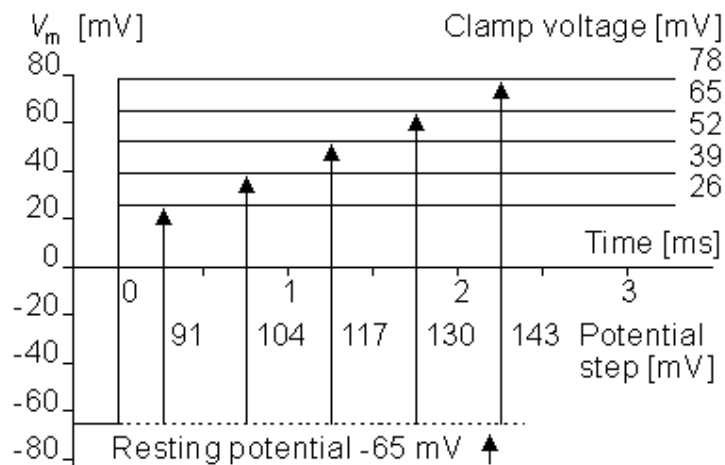


**Clamped voltage**



**Measured current**

# Hodgkin-Huxley: Measurement Protocols



## Protocols

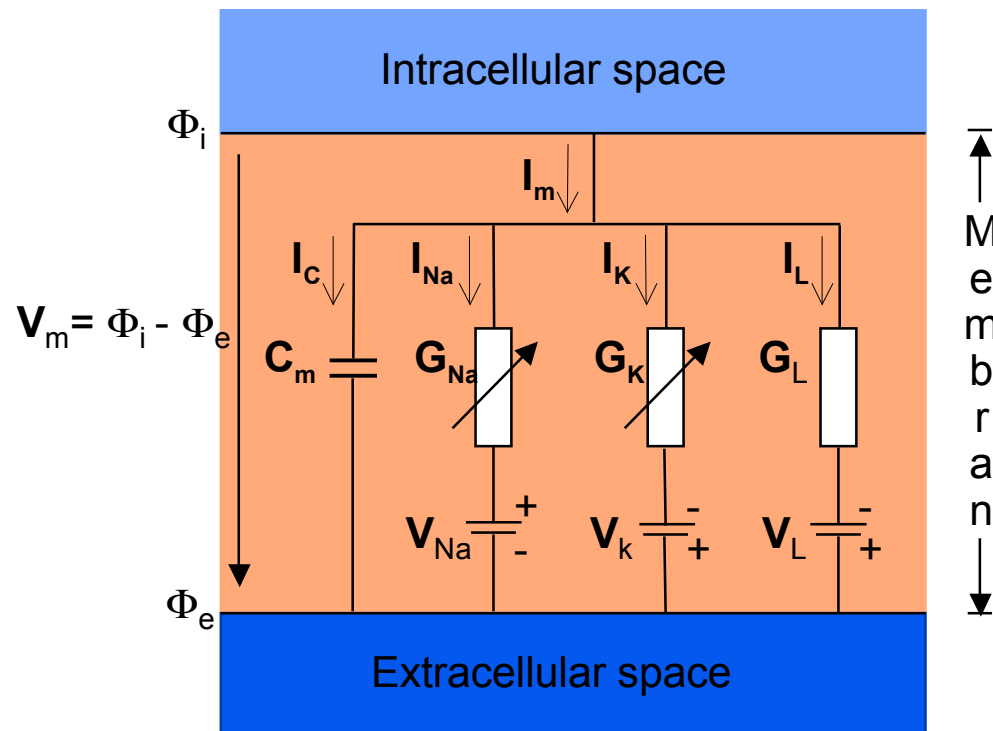
Variation of clamped voltages

Extraction of parameters from measured curves, e.g. peak current

Substitution of ions in intra- and extracellular space

(Curve fitting for modeling)

# Hodgkin-Huxley Model: Equivalent Circuit Diagram



$G_{Na}, G_K, G_L$   
Membrane conductivity of Na, K and other ions [S/cm<sup>2</sup>]

$I_{Na}, I_K, I_L$   
Currents of Na, K and other ions [mA/cm<sup>2</sup>]

$V_{Na}, V_K, V_L$   
Nernst voltages of Na, K and other ions [mV]

$C_m, I_m, V_m$   
Membrane capacitor [F/cm<sup>2</sup>], current [mA/cm<sup>2</sup>] and voltage [mV]

$$I_m = C_m \frac{dV_m}{dt} + (V_m - V_{Na})G_{Na} + (V_m - V_K)G_K + (V_m - V_L)G_L$$

# Hodgkin-Huxley Model: Principles

Ohm's law:

$$\cancel{G_{\text{Na}} = \frac{I_{\text{Na}}}{V_{\text{Na}}} \quad \cancel{G_{\text{K}} = \frac{I_{\text{K}}}{V_{\text{K}}} \quad \cancel{G_{\text{Na}} = \frac{I_{\text{L}}}{V_{\text{L}}}}$$

Nernst voltages for correction!

$$G_{\text{Na}} = \frac{I_{\text{Na}}}{V_{\text{m}} - V_{\text{Na}}} \quad G_{\text{K}} = \frac{I_{\text{K}}}{V_{\text{m}} - V_{\text{K}}} \quad G_{\text{Na}} = \frac{I_{\text{L}}}{V_{\text{m}} - V_{\text{L}}}$$



# Hodgkin-Huxley Model: Constants

Voltages are related to resting voltage  $V_r$   
 Conductivity and capacitance are related to membrane area

Relative Na voltage	$V_r - V_{Na}$	-115	mV	
Relative K voltage	$V_r - V_k$	12	mV	
Relative voltage of other ions	$V_r - V_L$	-10.6	mV	
Membrane capacitance	$C_m$	1	$\mu\text{F}/\text{cm}^2$	
Maximal conductivity of Na	$G_{Na \max}$	120	$\text{mS}/\text{cm}^2$	} All ion channels open
Maximal conductivity von K	$G_{K \max}$	36	$\text{mS}/\text{cm}^2$	
Conductivity for other ions	$G_L$	0.3	$\text{mS}/\text{cm}^2$	



# Hodgkin-Huxley Model: ODEs Describe Conductivities

$$\begin{aligned}
 G_{\text{Na}} &= G_{\text{Na max}} m^3 h & \frac{dm}{dt} &= \alpha_m(1-m) - \beta_m m & \left. \vphantom{\frac{dm}{dt}} \right\} & \text{Sodium current} \\
 & & \frac{dh}{dt} &= \alpha_h(1-h) - \beta_h h & & \\
 G_{\text{K}} &= G_{\text{K max}} n^4 & \frac{dn}{dt} &= \alpha_n(1-n) - \beta_n n & \left. \vphantom{\frac{dn}{dt}} \right\} & \text{Potassium current} \\
 G_{\text{L}} &= \text{const} & & & \left. \vphantom{\frac{dn}{dt}} \right\} & \text{Current by other ions}
 \end{aligned}$$

$$\alpha_m = \frac{0.1(25 - V')}{e^{0.1(25 - V')} - 1} \frac{1}{\text{ms}}$$

$$\alpha_h = \frac{0.07}{e^{V'/20}} \frac{1}{\text{ms}}$$

$$\alpha_n = \frac{0.01(10 - V')}{e^{0.1(10 - V')} - 1} \frac{1}{\text{ms}}$$

$$\beta_m = \frac{4}{e^{V'/18}} \frac{1}{\text{ms}}$$

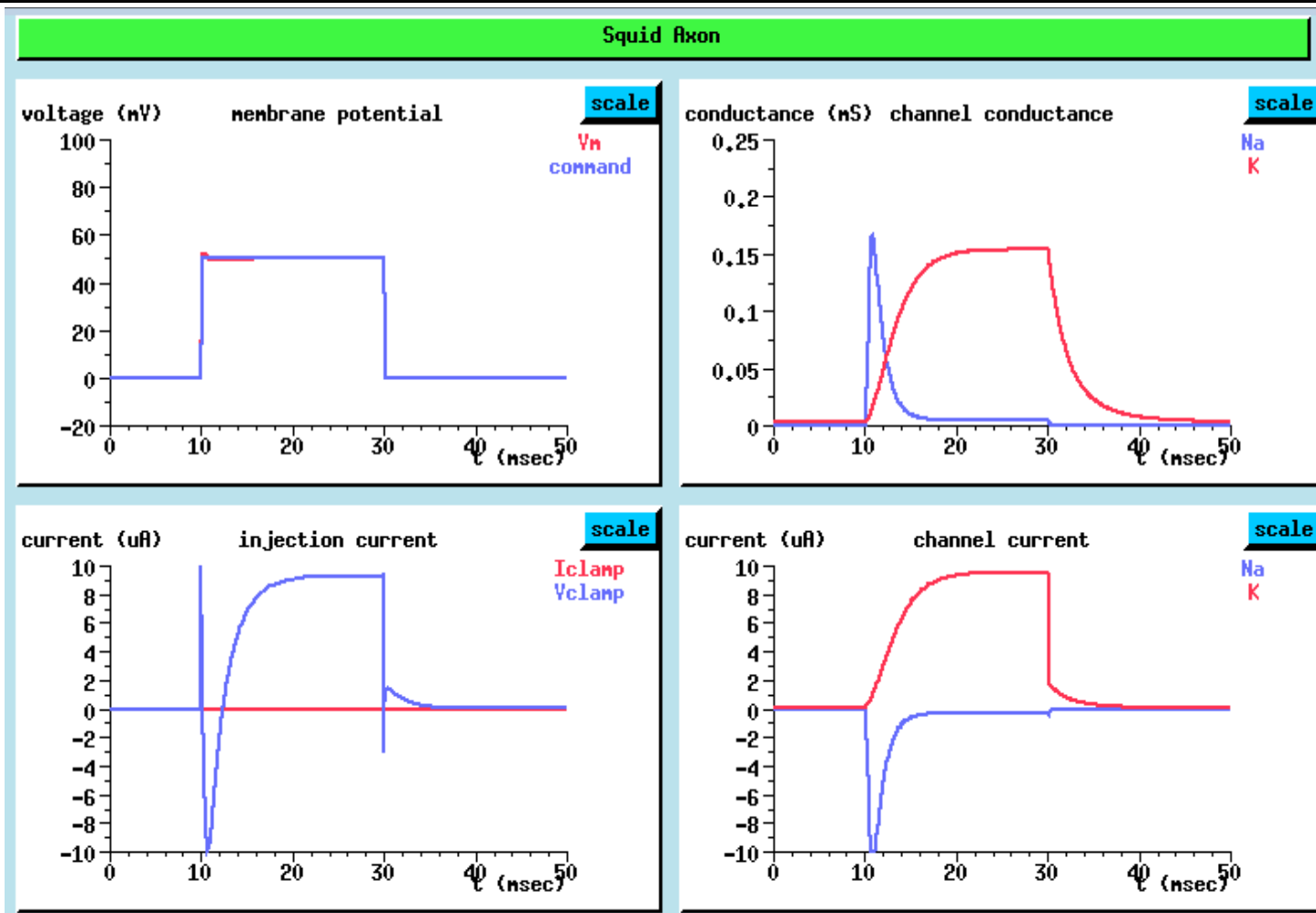
$$\beta_h = \frac{1}{e^{0.1(30 - V')} + 1} \frac{1}{\text{ms}}$$

$$\beta_n = \frac{0.125}{e^{V'/80}} \frac{1}{\text{ms}}$$

Voltage and time-dependent



# Hodgkin-Huxley Model: Simulation of Voltage Clamp Measurements

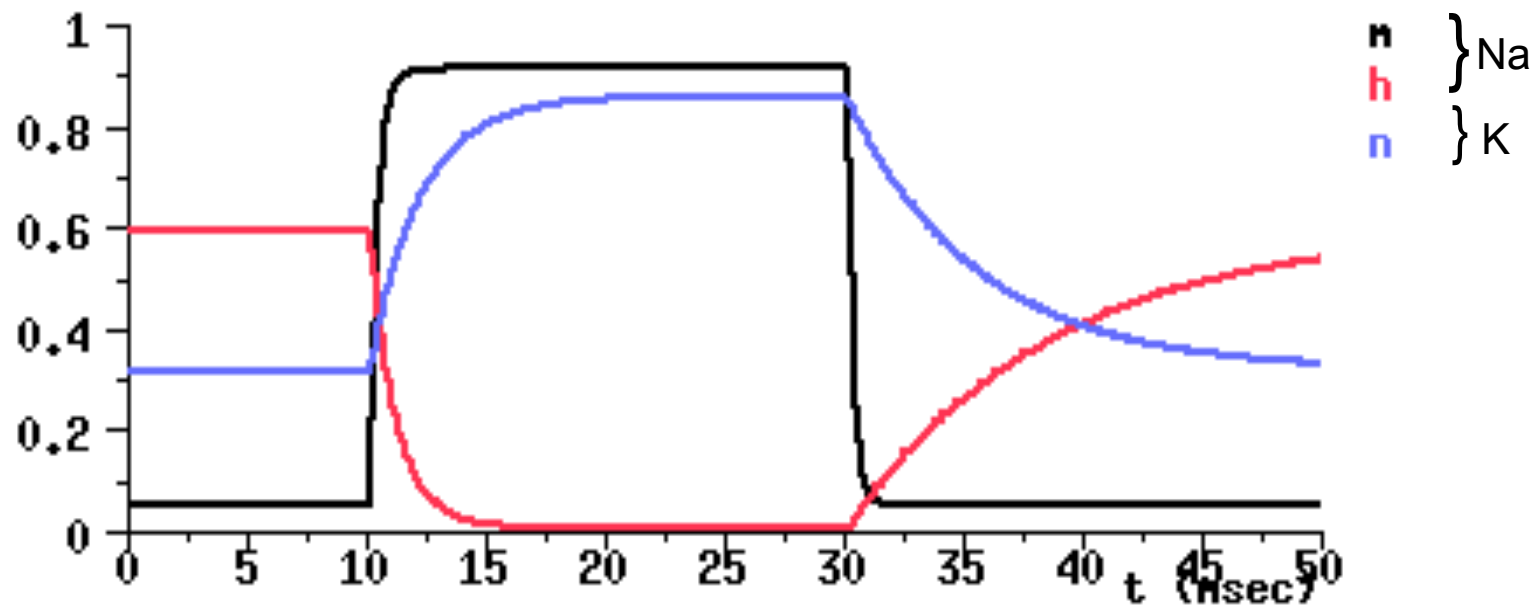


<http://www.bbb.caltech.edu/GENESIS>

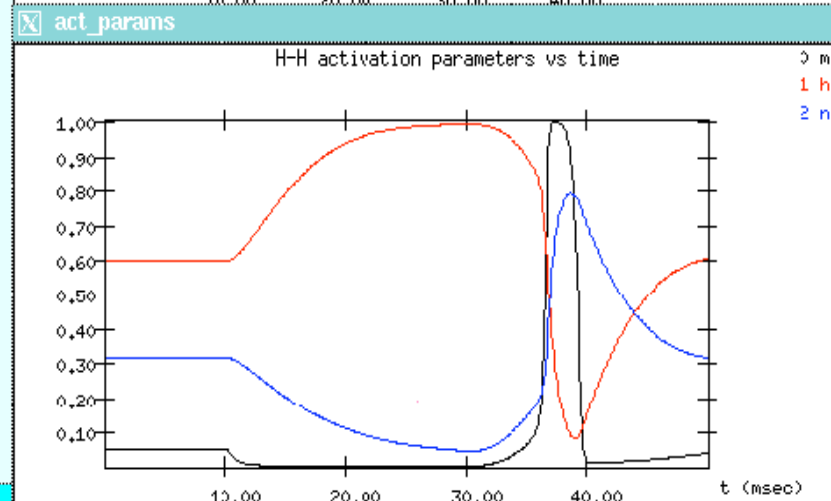
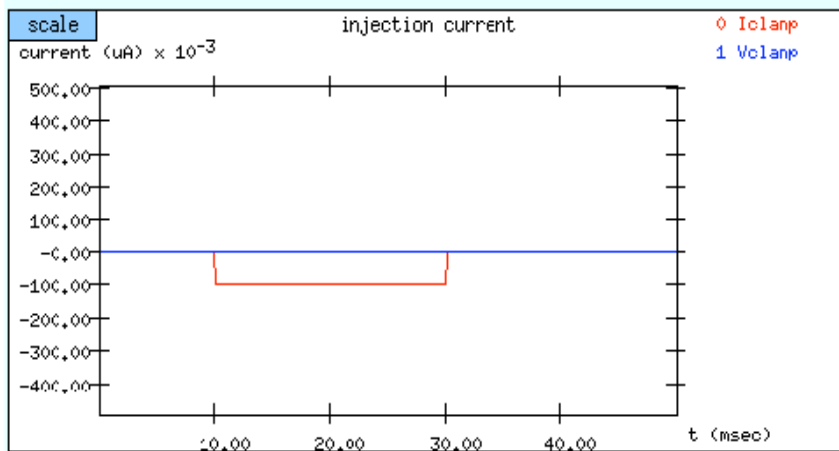
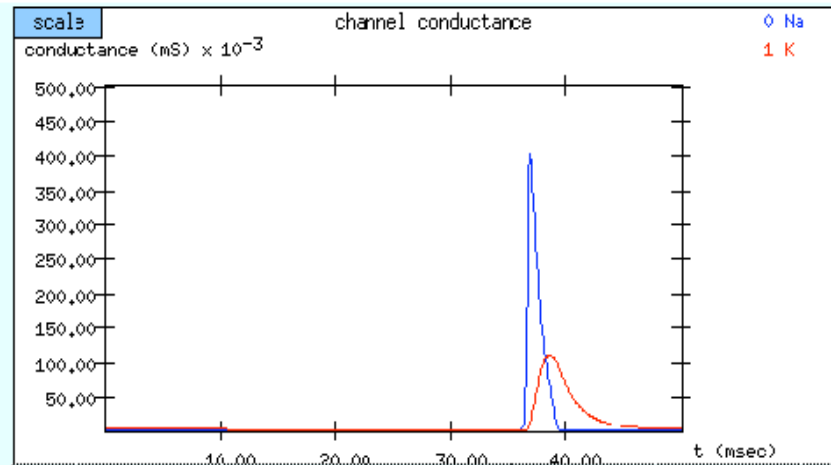
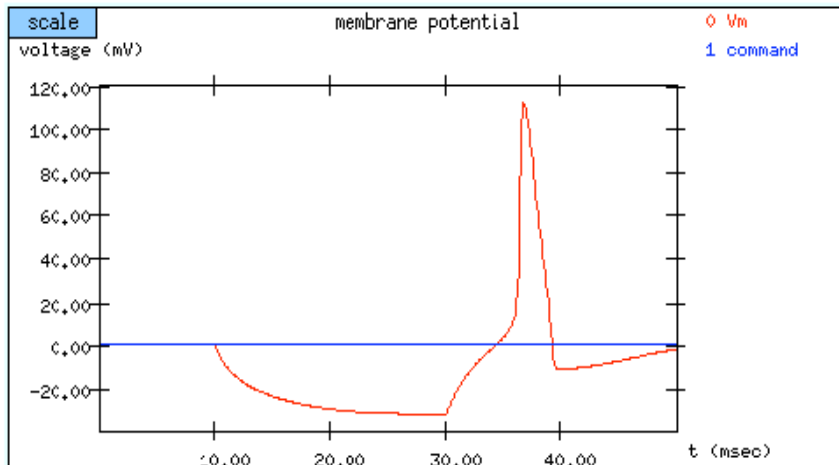
# Hodgkin-Huxley Model: Simulation of Voltage Clamp Measurements

$$G_{\text{Na}} = G_{\text{Na,max}} m^3 h$$

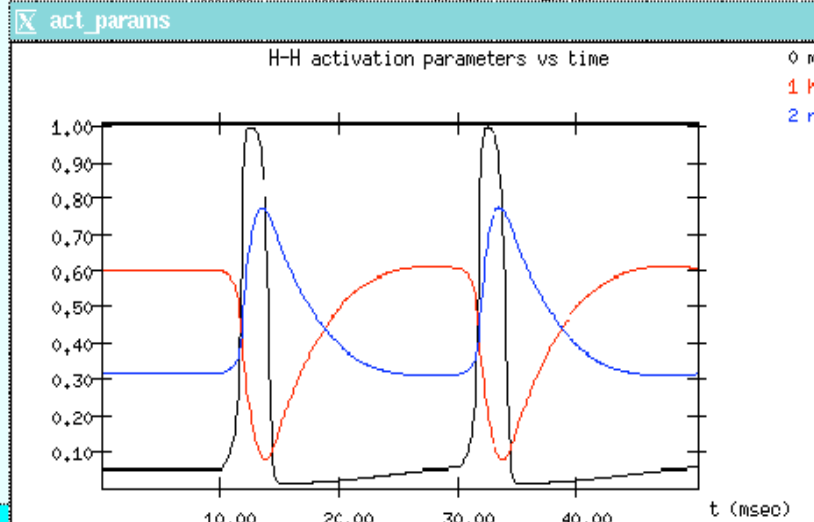
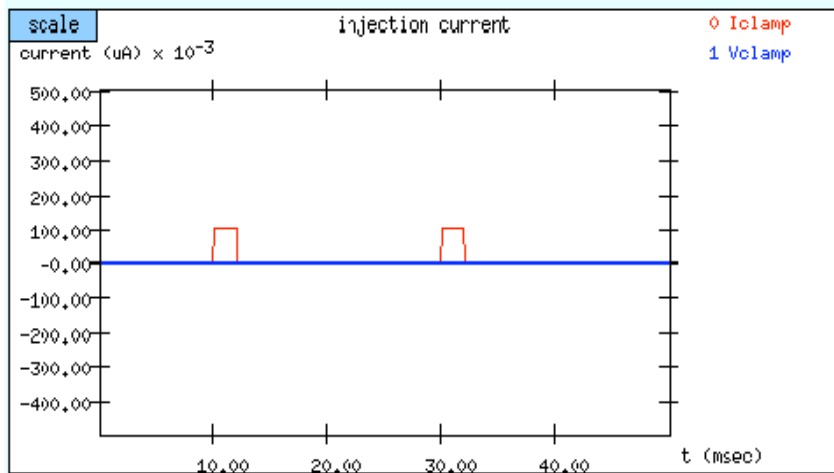
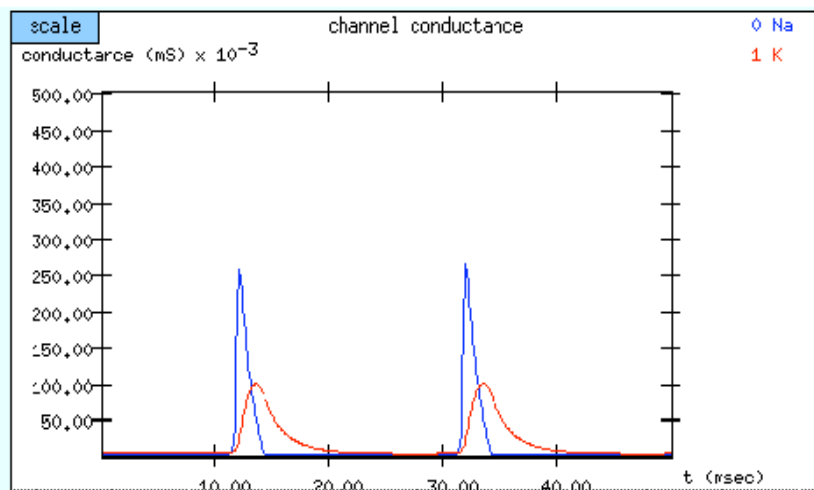
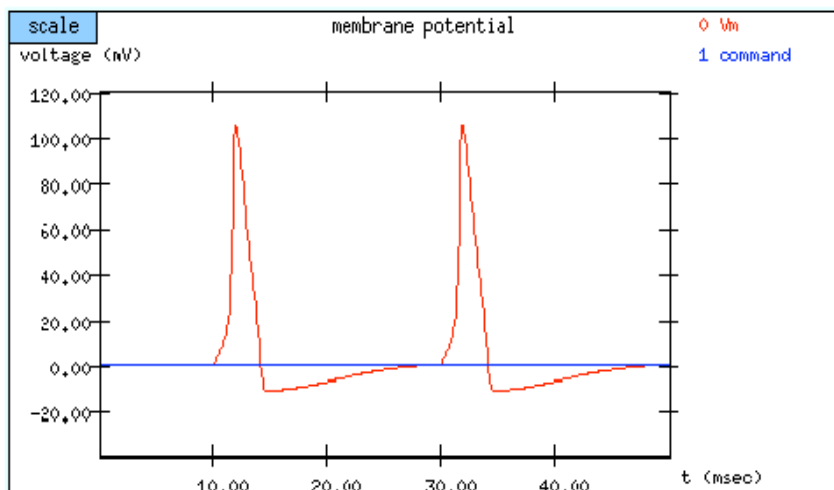
$$G_{\text{K}} = G_{\text{K,max}} n^4$$



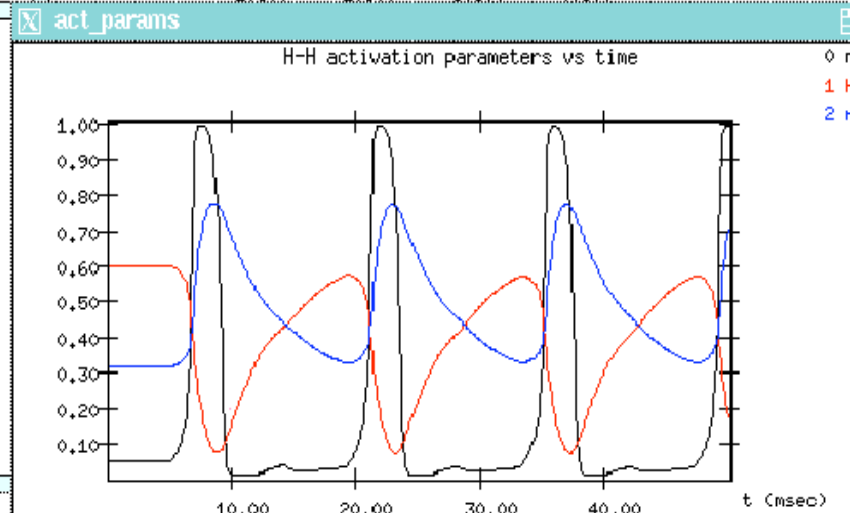
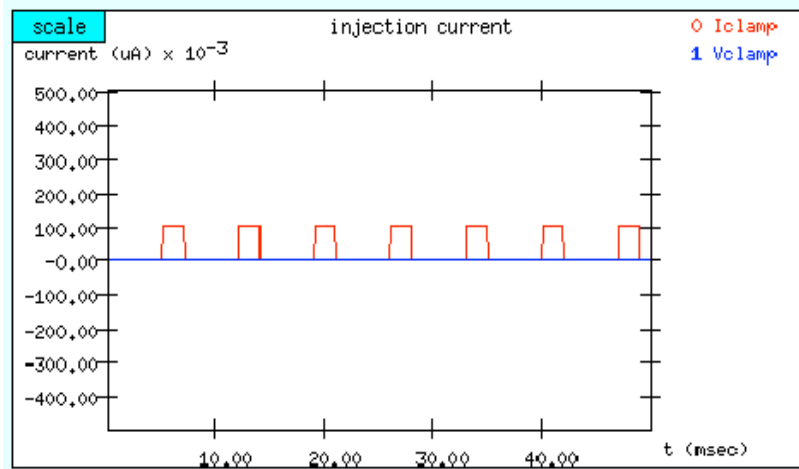
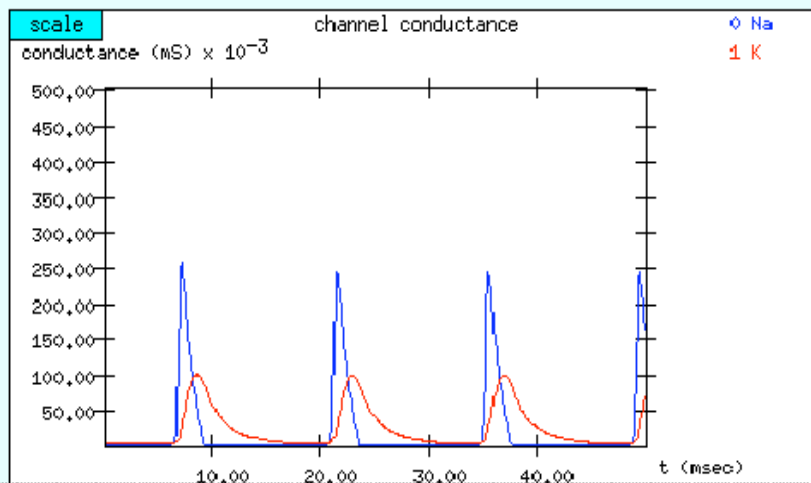
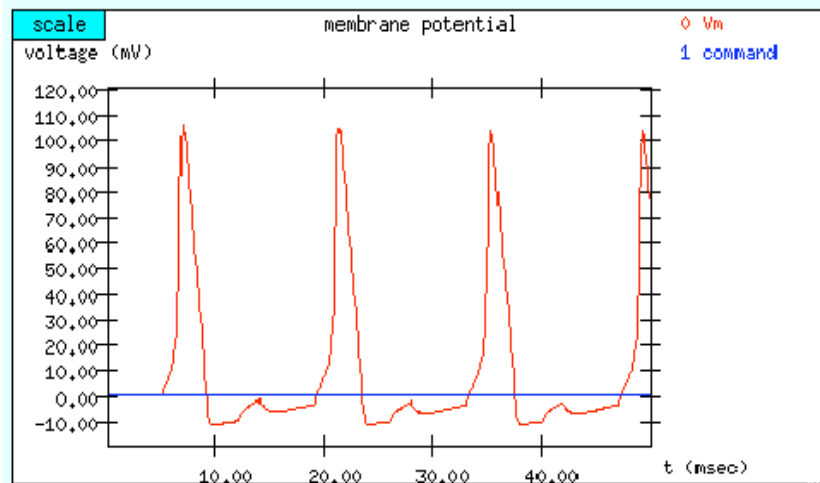
# Hodgkin-Huxley Model: Activation by Hyperpolarization



# Hodgkin-Huxley Model: Stimulus After Refractory Period



# Hodgkin-Huxley: Stimulus During Refractory Period



## Group Work

Sodium channels allow fast upstroke of action potentials of neurons and myocytes.

Speculate why fast voltage-dependent inactivation of these channels might be important.

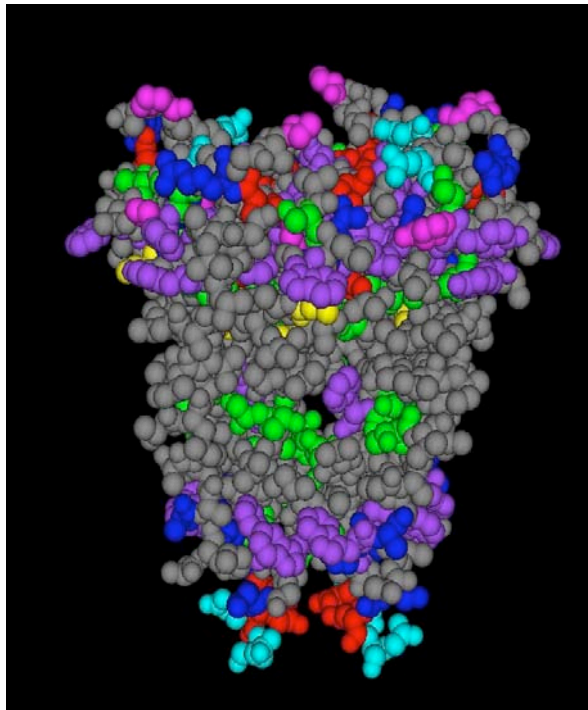


# Molecular Structure of Ion Channels

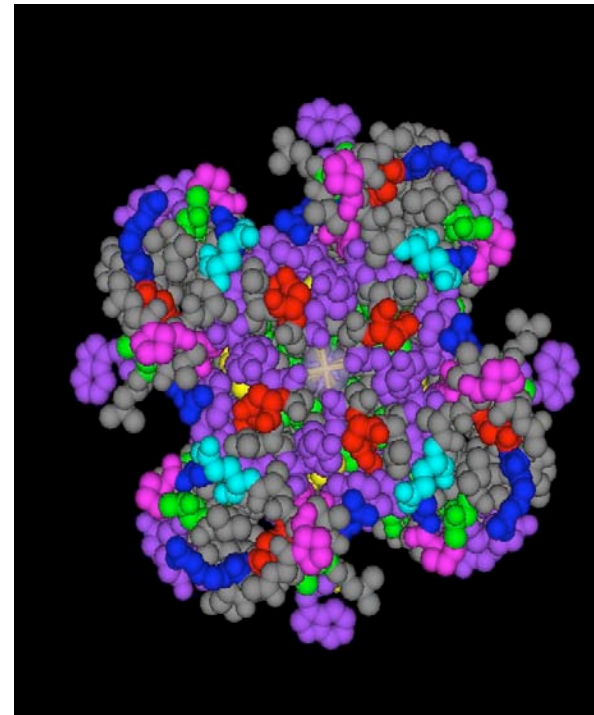
Transmembrane proteins: connexons, **ion channels**, pumps and exchangers

Example: Molecular structure of potassium channel Kcsa of bacterium streptomyces lividans, color-coded amino acids  
Structure data from Molecular Modeling Database, NIH, USA

~ 6 nm

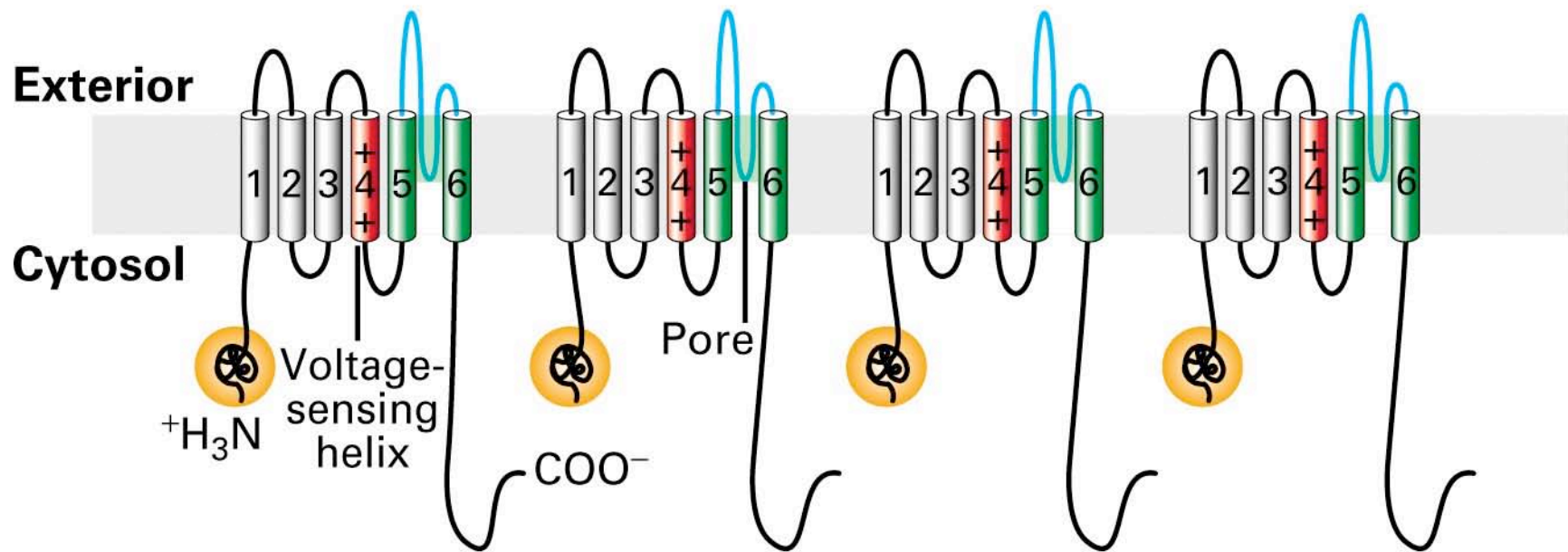


side

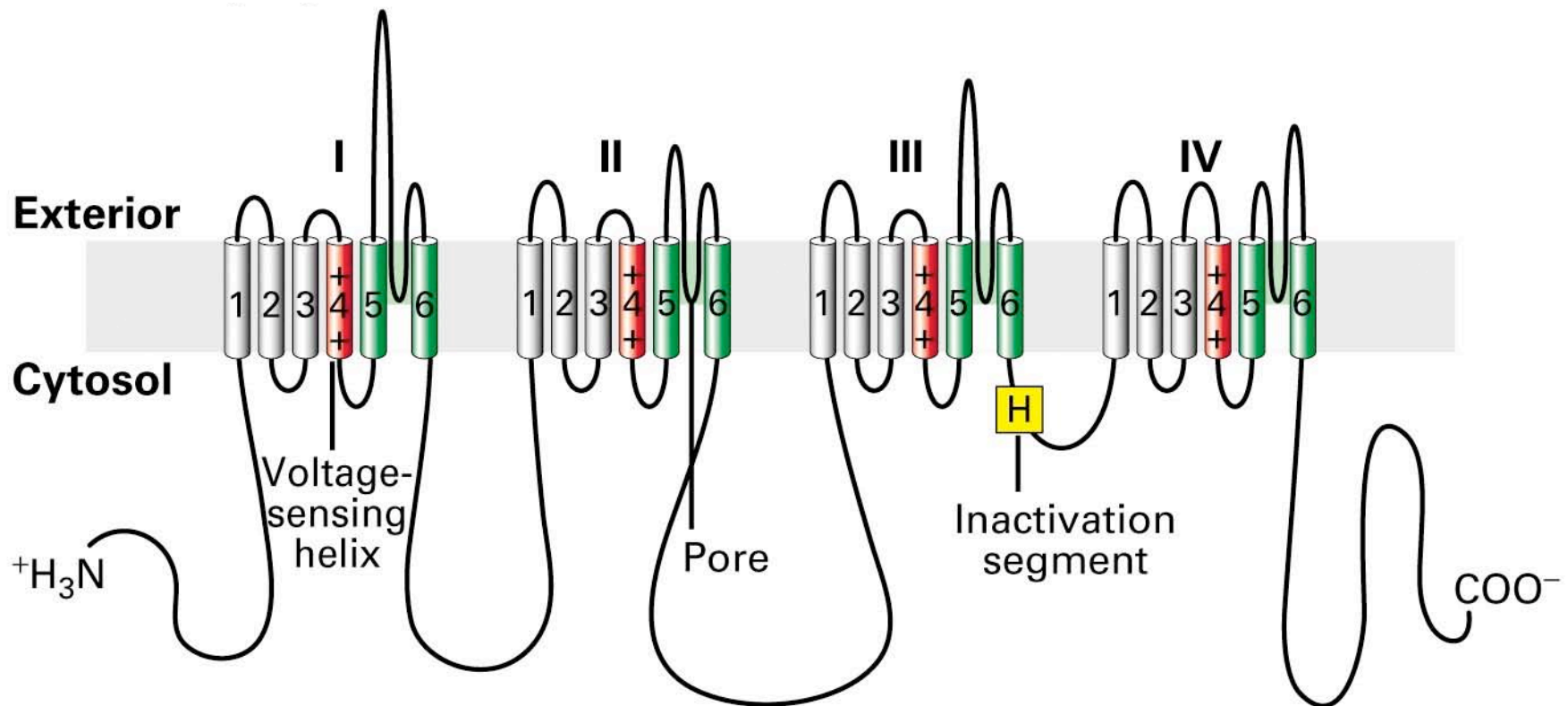


from  
top

# Schematic Depiction of Voltage-Gated K<sup>+</sup> Channel (Tetramer)



## Schematic Depiction of Voltage-Gated Na<sup>+</sup>/Ca<sup>2+</sup> Channel (Monomer)



# Experimental Studies: Patch Clamp Techniques

Measurement technique developed by  
Neher, Sakmann et al.  
(published 1976, Nobel prize 1991)

## Micropipettes

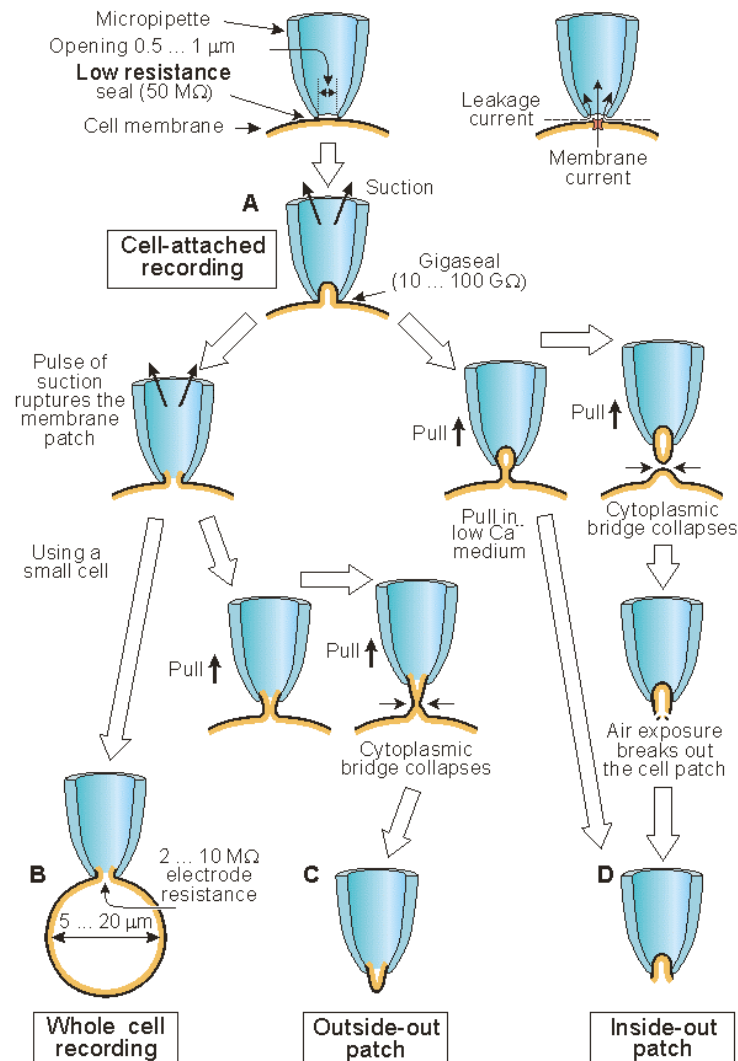
- heat polished fluid filled glass pipette
- diameter of opening: 0.5-1  $\mu\text{m}$

## Major configurations

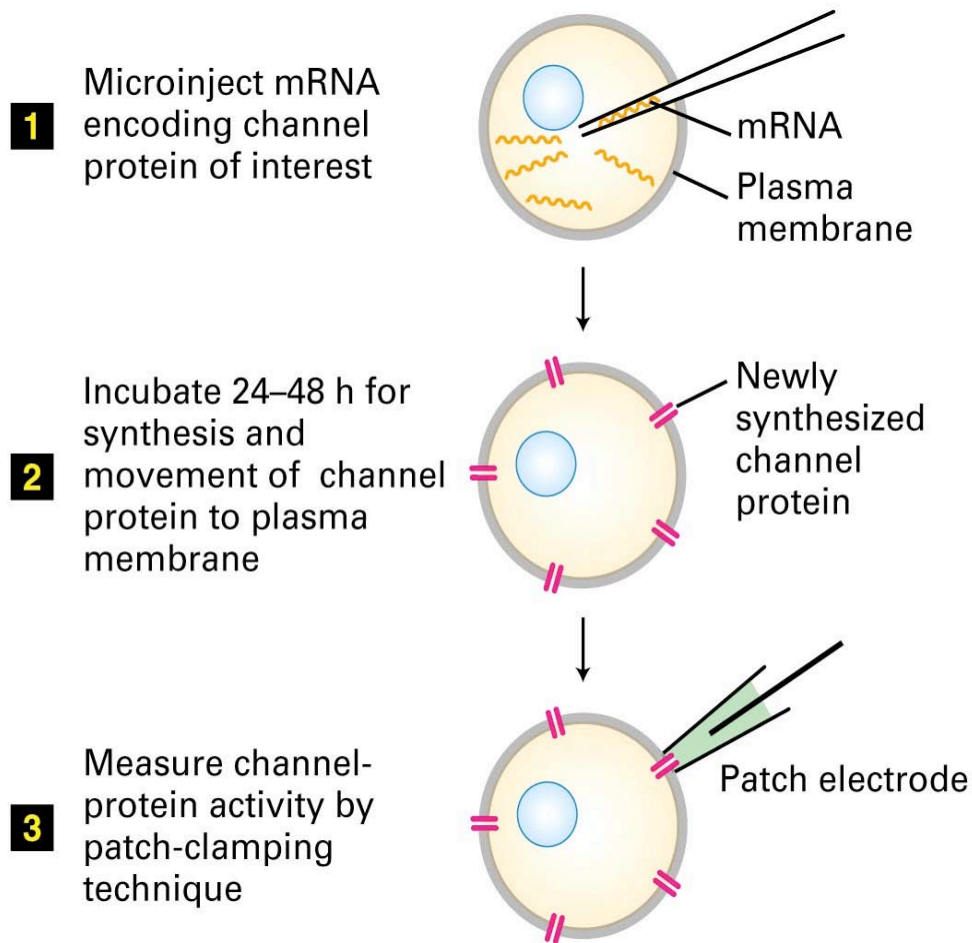
- Cell attached recording
- Whole cell recording
- Outside-out patch
- Inside-out patch

Measurement of single ion channels  
possible!

Commonly, signals have small  
signal-to-noise ratios

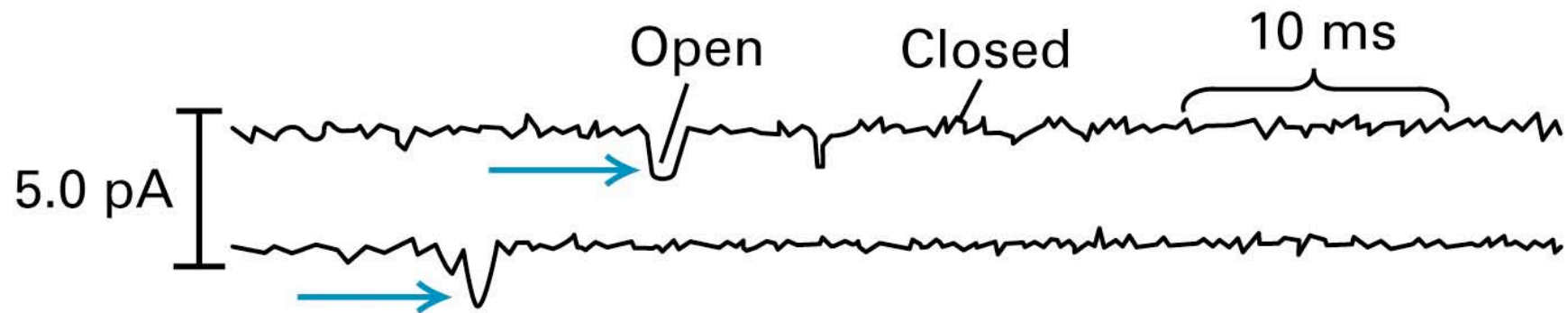


# Channel Characterization in Oocyte Expression Array



(Lodish et al., Molecular Cell Biology, Fig. 7-19, 2004)

## Currents Through Single Ion Channel

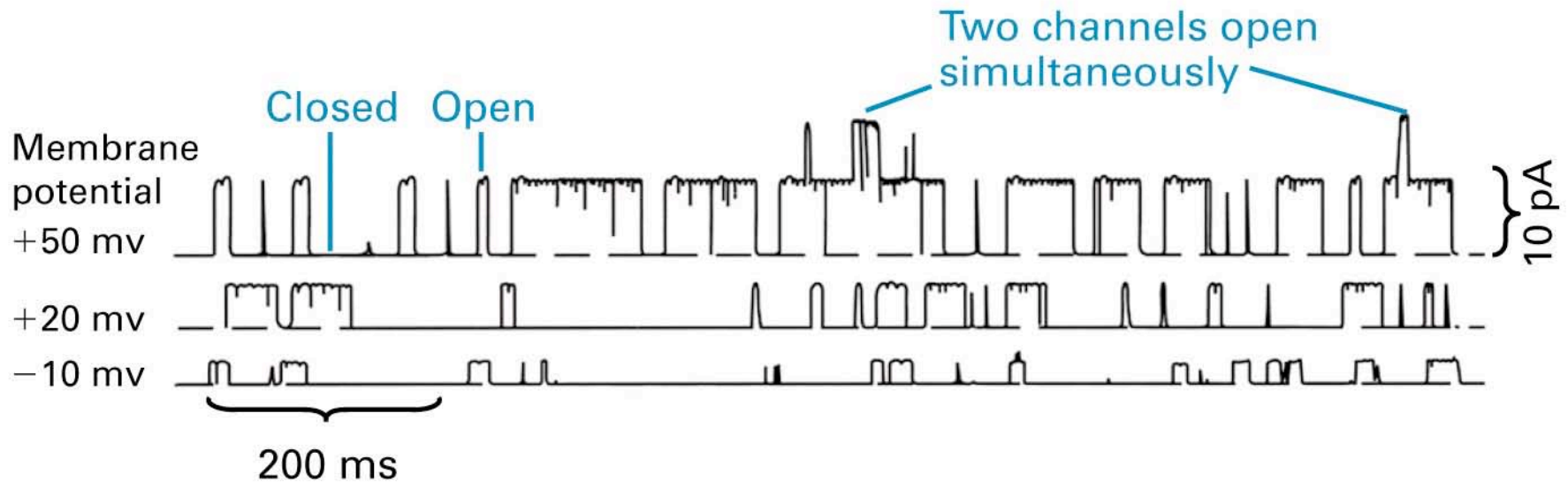


Current traces of patch with single sodium channel

Average current per channel: 1.6 pA ~ 9900 ions/ms

Inside-out patch

# Currents Through Ion Channels



Current traces of patch with 2 potassium channels at different voltages

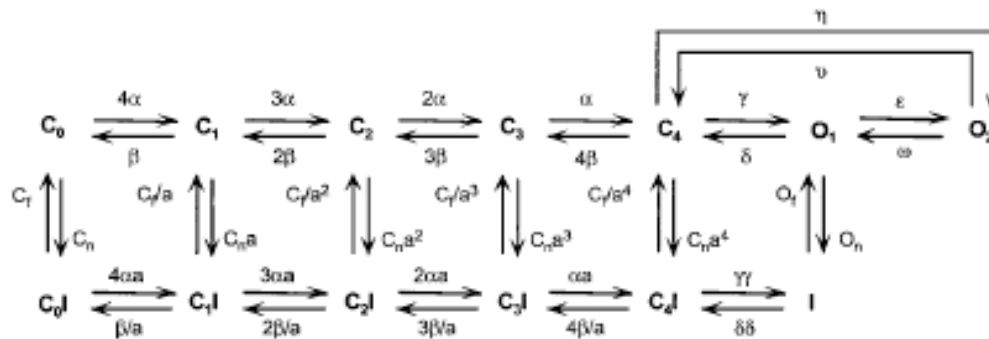
Transmembrane voltages determine

- open probability
- open time
- current amplitude

# Markov Modeling of Ion Channels and Mutations

## Markov models allow

- reconstruction of single channel behavior
- to be based upon thermodynamic principals
- assignment of physical meaning to rate constants



**Example:** State diagram of cardiac sodium channel model

O: Open, I: Inactivated, C: Closed

(Irvine et al. Biophys J. 1999)

- Markov models consist of sets of 1st order ODEs
- Commonly, one channel description of a “traditional” Hodgkin-Huxley type cell model is substituted by an appropriate Markov model
- Recently, the inclusion of Markov models in newly developed cell models increased





## 2-State Markov Model

$$\frac{dO}{dt} = \alpha C - \beta O$$

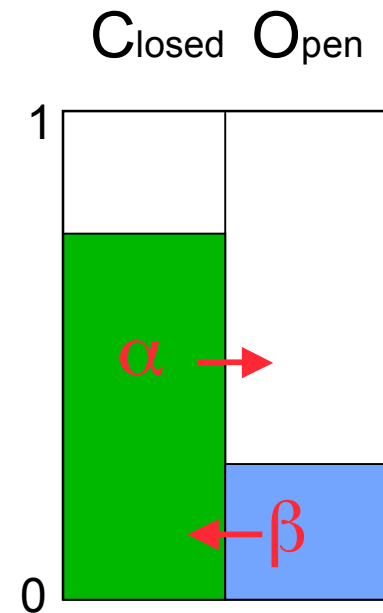
$$\frac{dC}{dt} = \beta O - \alpha C$$

O : Probability of channel is in open state

C : Probability of channel is in closed state

$\alpha, \beta$  : Rate coefficients.

Function of e.g.  $V_m$  and ion concentration



## Exemplary Rate Coefficient Functions

$$\alpha = \alpha_0$$

Constant

$$\alpha = \alpha_0 V_m + a$$

Linear

$$\alpha = \alpha_0 e^{V_m/a}$$

Exponential

$$\alpha = \frac{\alpha_0}{e^{-(V_m - V_a)/a} + 1}$$

Sigmoid

$$\alpha = \alpha_0 \frac{V_m - V_a}{e^{-(V_m - V_a)/a} - 1}$$

Linear for extreme case

$\alpha_0, V_a, a$  : Parameters

$V_m$  : Membrane voltage

## Calculation of Membrane Currents: Variants

$$I_{\text{chan}} = N G O (V_m - E_{\text{ion}})$$

Nernst approach

$$I_{\text{ion}} = P z^2 \frac{F^2 V_m}{RT} \frac{[\text{ion}]_i - [\text{ion}]_o e^{-z F V_m / RT}}{1 - e^{-z F V_m / RT}}$$

Goldman – Hodgkin – Katz  
current equations

G: Conductivity of single channel

O: Open probability of channels

N: Number of channels

$V_m$ : Membrane voltage

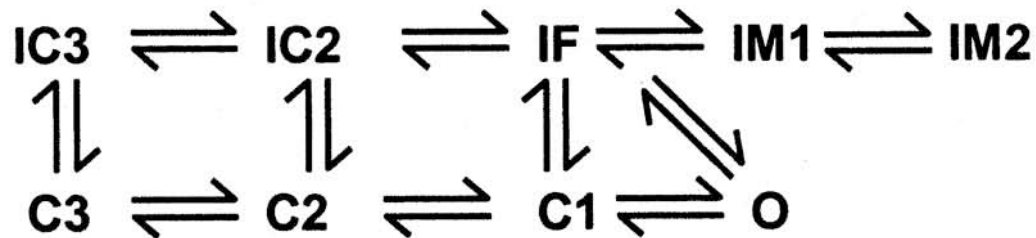
P: Membrane permeability for ion

$[\text{ion}]_i$ ,  $[\text{ion}]_o$ : Concentration of ion in intra- and extracellular space

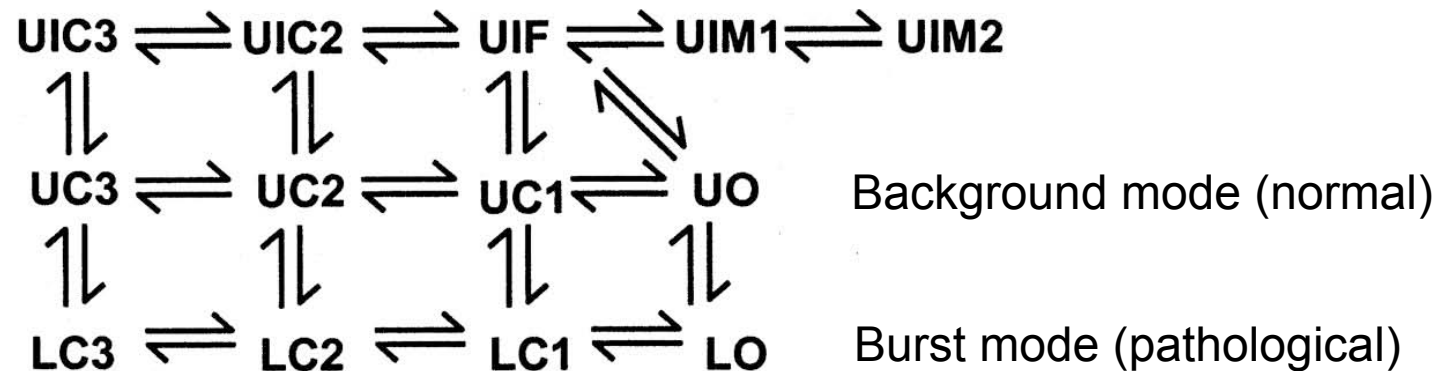


# Markov Models for WT and 1795insD Cardiac Na Channels

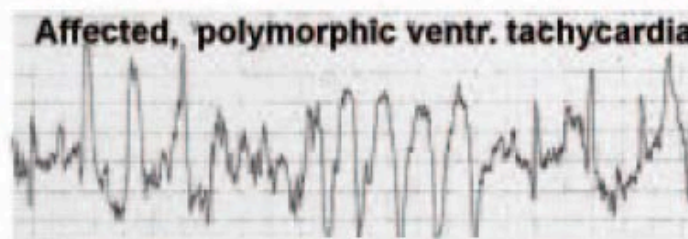
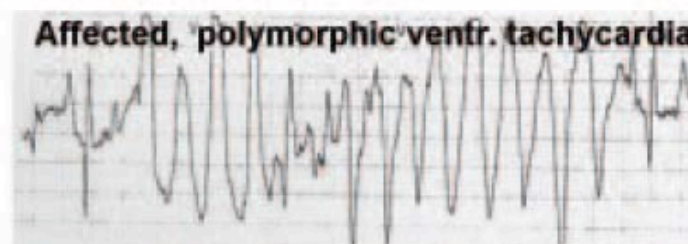
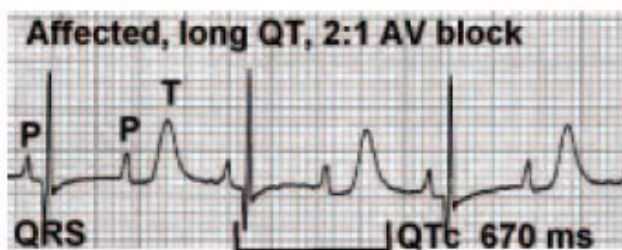
Wild-type Na channel



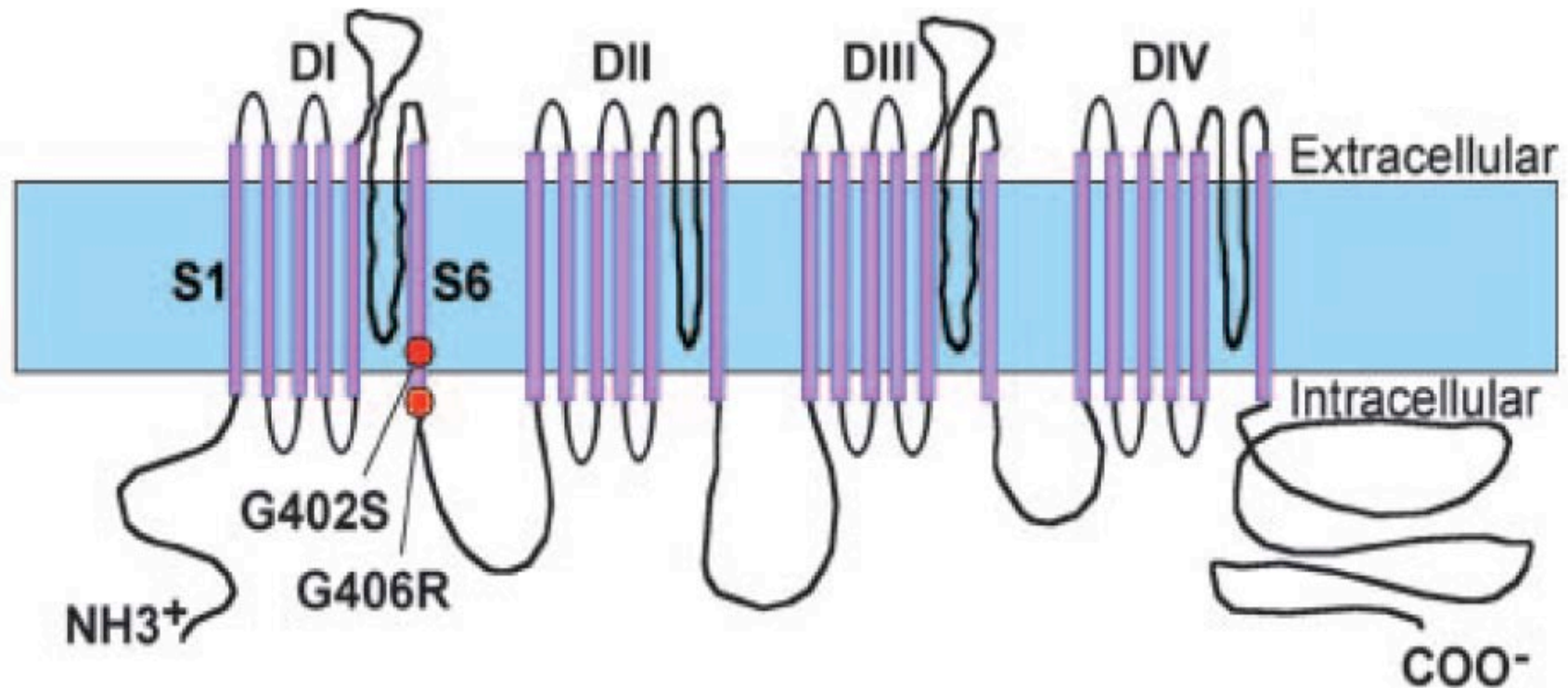
1795insD Na channel



# Genetic Disease: Timothy Syndrome

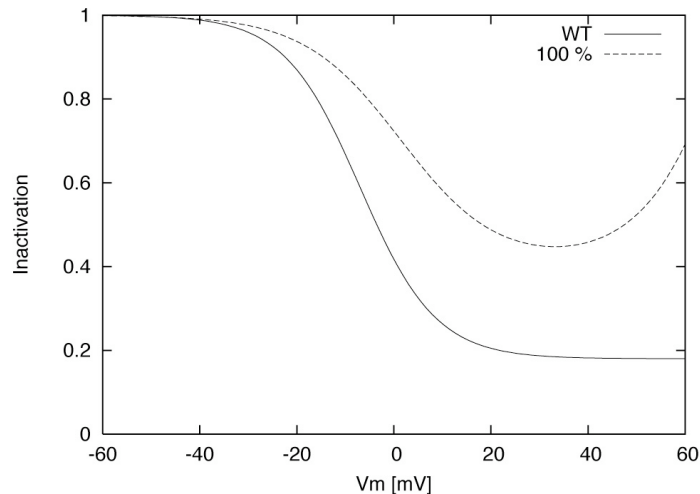


# Topology of Ion Channel Protein Ca<sub>v</sub>1.2



“De novo” mutations { G402S      Glycine → Serine  
                                   G406R      Glycine → Arginine

# Modeling of Timothy Syndrome



## Channel Modeling

Differences of steady state inactivation between wild type (WT) and mutated channels

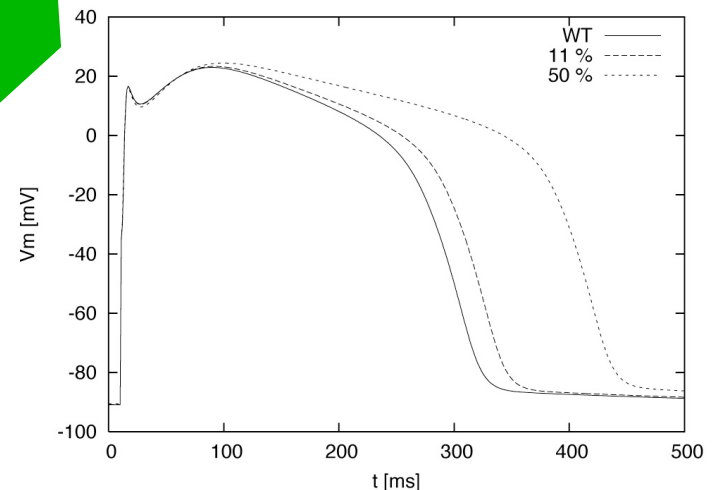
Numerical optimization

Integration in Myocyte Model

## Prediction of course of transmembrane voltage in myocyte

Changes dependent on % of channels with mutation

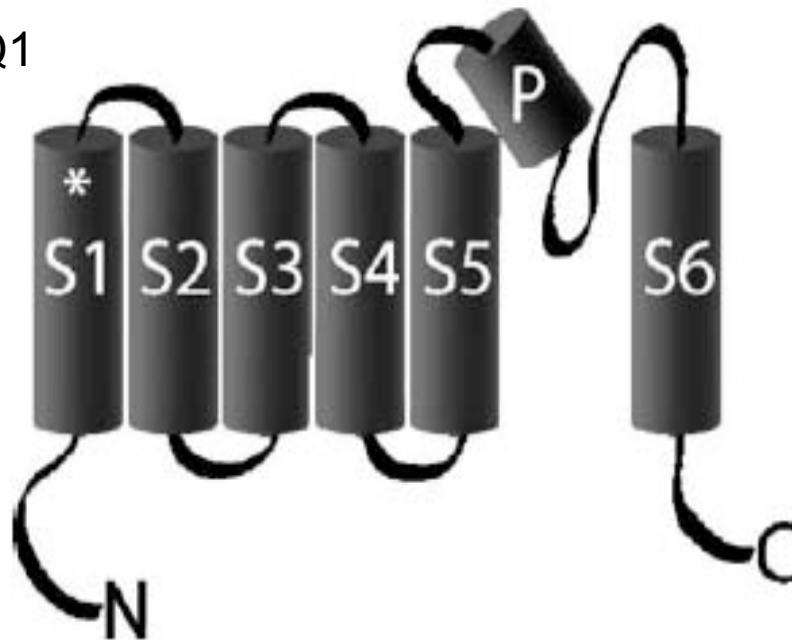
Significant increase of action potential duration (and intracellular calcium concentrations)



# Genetic Disease: Mutation of KCNQ1

Slow Inward Rectifying Potassium Current  $I_{Ks}$   $\left\{ \begin{array}{l} \text{KCNQ1} \\ \text{KCNE1} \end{array} \right.$

KCNQ1



## Mutations

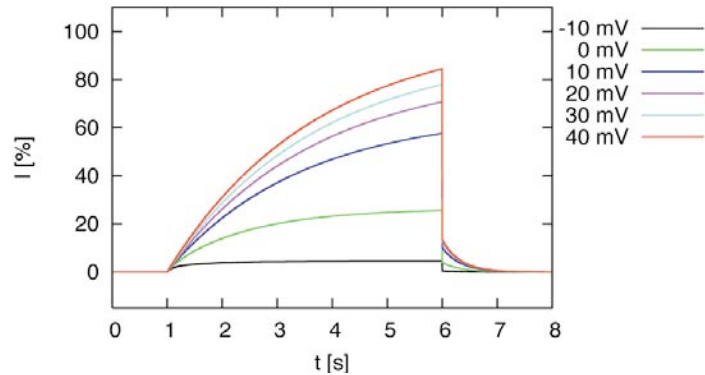
- **S140G**  
Serine  $\rightarrow$  Glycine  
found in family with hereditary atrial fibrillation  
(Chen et al., Science, 2003)
- **V141M**  
Valine  $\rightarrow$  Methionine  
found in new born child with atrial fibrillation and short QT syndrome “de novo”  
(Kong et al., Cardiovasc Res, 2005)

\* Location of Mutation S4: Voltage sensing subunit

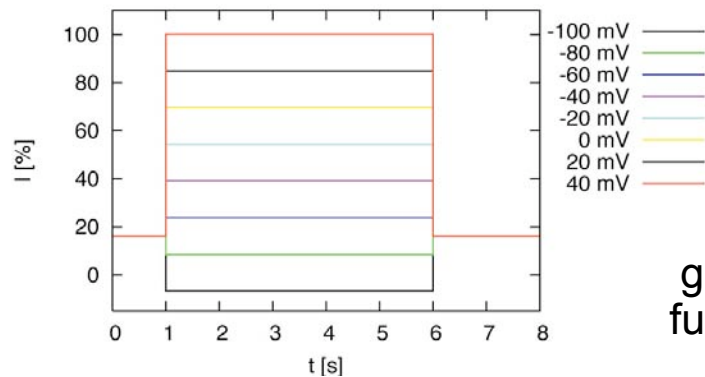


# Modeled Channel Data

## WT KCNQ1 + KCNE1

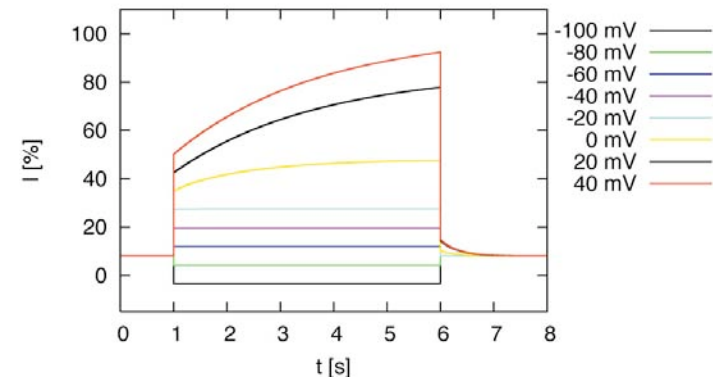


## KCNQ1 (S140G or V141M) + KCNE1



gain of function!

## 50 % WT / 50 % mutation



(Kong et al., Cardiovasc Res, 2005)

# Ordinary Differential Equations (ODEs)

ODEs of n-th order can be reduced to set of 1st order ODEs

2nd order ODE

$$\frac{\partial^2 u}{\partial x^2} + q(x) \frac{\partial u}{\partial x} = r(x)$$

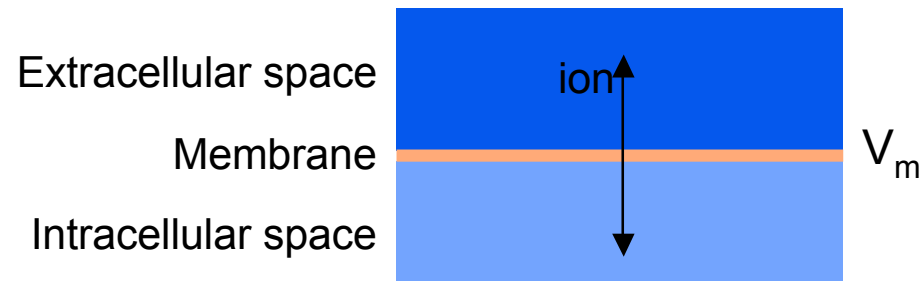
Rewrite

System of 1st order ODEs

$$\frac{\partial u}{\partial x} = z(x)$$

$$\frac{\partial z}{\partial x} = r(x) - q(x)z(x)$$

# 1st Order ODE for Describing Transmembrane Voltage



$$I_m = I_i + C_m \frac{d}{dt} V_m$$

$I_i$  : Injected current [A]

$I_m$  : Current through membrane [A]

$C_m$  : Membrane capacitor [F]

$V_m$  : Membrane voltage [V]

# Numerical Solution of ODEs

## Procedure

Discretization:  $\frac{\partial u}{\partial x} \rightarrow \frac{\Delta u}{\Delta x}$

Choose appropriate step length  $\Delta x$ : Distance between  $x_n$  and  $x_{n+1}$   
Determining factor for numerical error

## Numerical Methods

- Euler Method
- Runge-Kutta Method 2. Order
- Runge-Kutta Method 4. Order
- Richardson-Extrapolation, Bulirsch-Stoer Method
- Predictor-Corrector Methods
- ...

# Euler Method

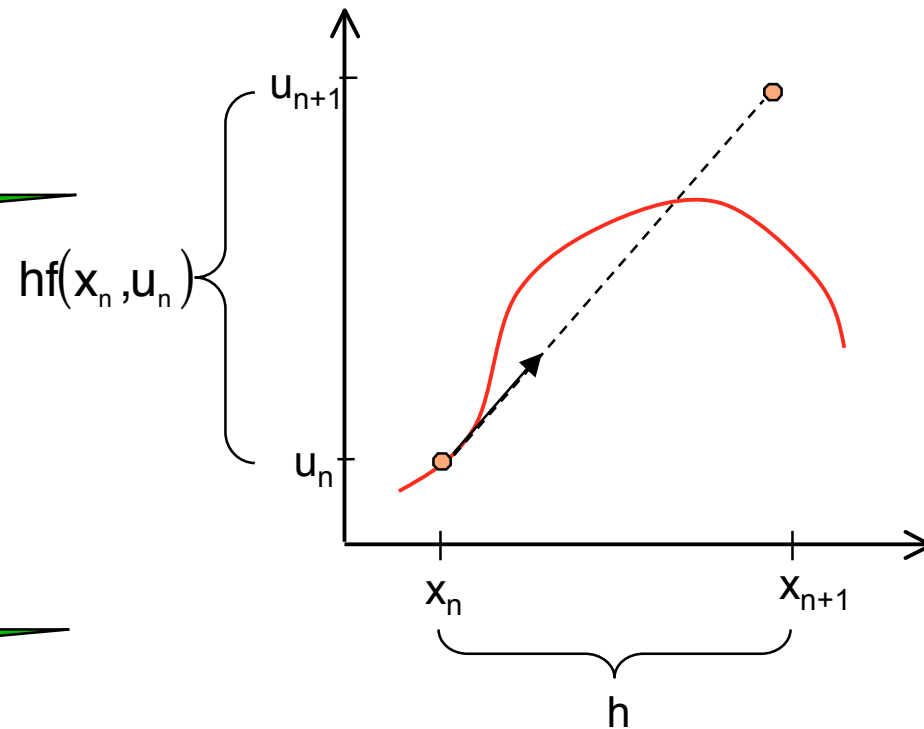
$$\frac{\partial u}{\partial x} = f(x, u)$$

**Finite Difference Approximation**

$$\frac{u_{n+1} - u_n}{x_{n+1} - x_n} = f(x_n, u_n)$$

**Rewriting**

$$u_{n+1} = u_n + h f(x_n, u_n)$$



# Euler Method: Example

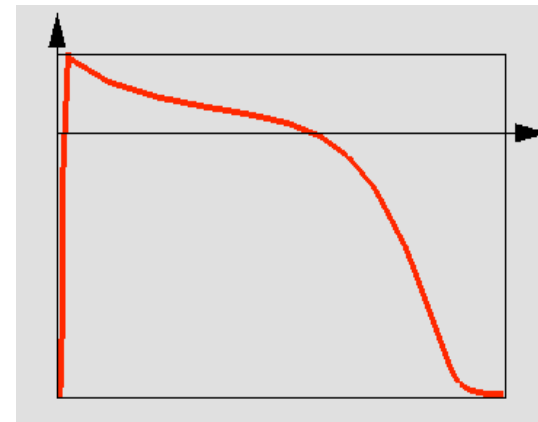
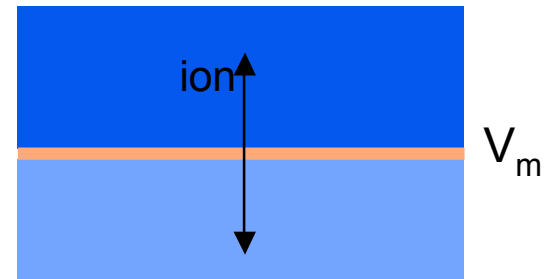
$$\frac{dV_m}{dt} = I_{stim}(t) - \frac{1}{C_m} I_{ion}(t, V_m)$$

Finite Difference Approximation

$$\frac{V_{n+1} - V_n}{t_{n+1} - t_n} = I_{stim}(t_n) - \frac{1}{C_m} I_{ion}(t_n, V_n)$$

Rewrite

$$V_{n+1} = V_n + \Delta t \left( I_{stim}(t_n) - \frac{1}{C_m} I_{ion}(t_n, V_n) \right)$$



# Runge-Kutta Method 2nd Order

$$\frac{\partial u}{\partial x} = f(x, u)$$

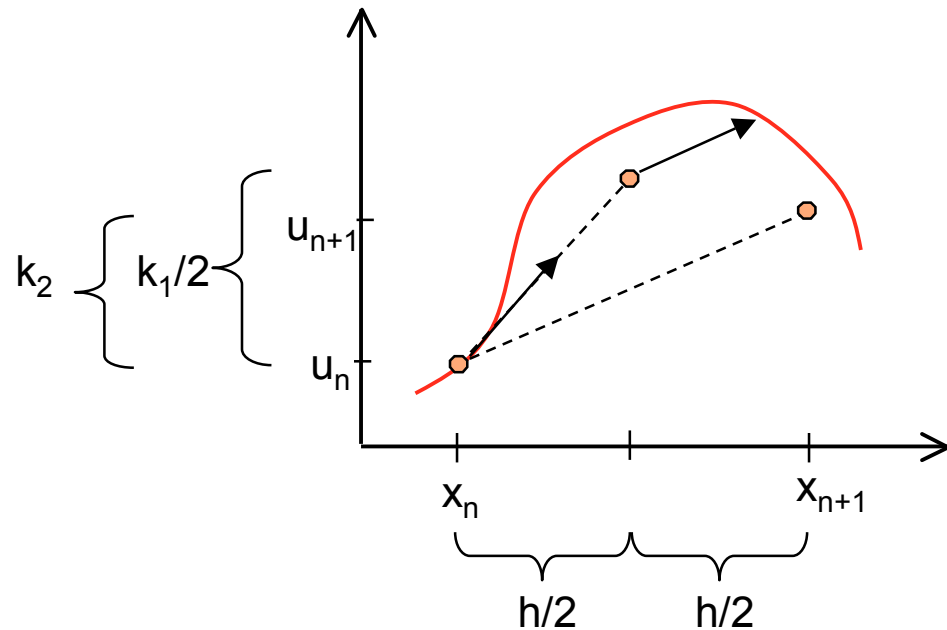
**Discretization**

$$k_1 = hf(x, u_n)$$

$$k_2 = hf\left(x_n + \frac{1}{2}h, u_n + \frac{1}{2}k_1\right)$$

**Step**

$$u_{n+1} = u_n + k_2$$



# Runge-Kutta Method 2nd Order: Example

$$\frac{dV_m}{dt} = I_{stim}(t) - \frac{1}{C_m} I_{ion}(t, V_m)$$

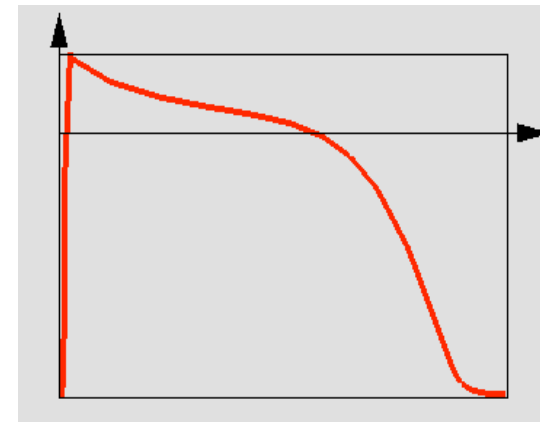
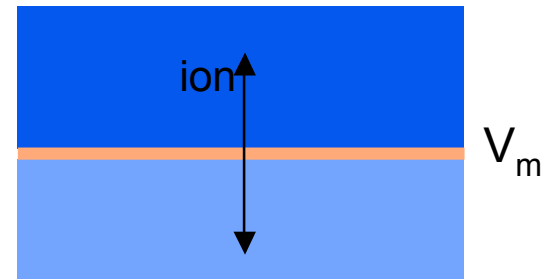
**Discretization**

$$k_1 = \Delta t \left( I_{stim}(t_n) - \frac{1}{C_m} I_{ion}(t_n, V_n) \right)$$

$$k_2 = \Delta t \left( I_{stim}\left(t_n + \frac{h}{2}\right) - \frac{1}{C_m} I_{ion}\left(t_n + \frac{h}{2}, V_n + \frac{k_1}{2}\right) \right)$$

**Step**

$$V_{n+1} = V_n + k_2$$





# Runge-Kutta Method 4th Order

$$\frac{\partial u}{\partial x} = f(x, u)$$

Discretization

$$k_1 = hf(x_n, u_n)$$

$$k_2 = hf\left(x_n + \frac{1}{2}h, u_n + \frac{1}{2}k_1\right)$$

$$k_3 = hf\left(x_n + \frac{1}{2}h, u_n + \frac{1}{2}k_2\right)$$

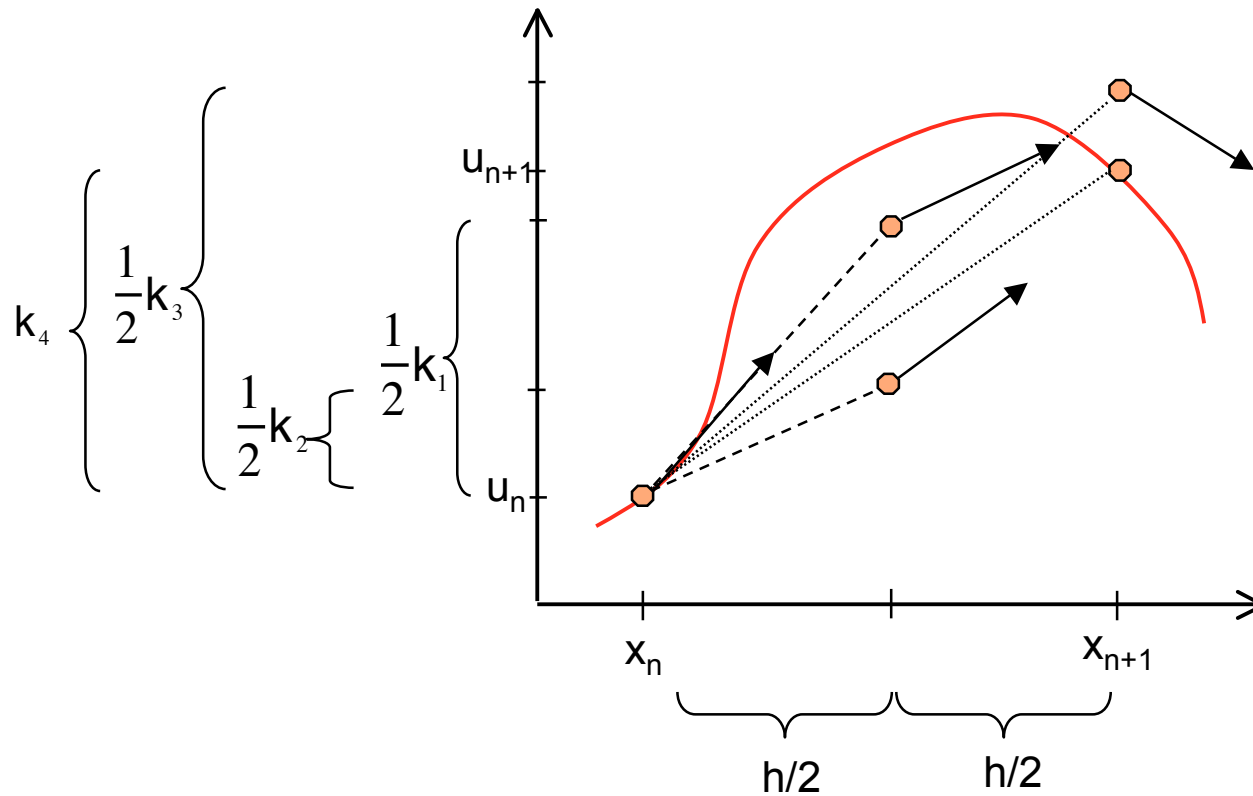
$$k_4 = hf(x_n + h, u_n + k_3)$$

Step

$$u_{n+1} = u_n + \frac{k_1}{6} + \frac{k_2}{3} + \frac{k_3}{3} + \frac{k_4}{6}$$



# Runge-Kutta Method 4th Order: Example



## Group Work

Solve manually applying some steps of the Euler method:

$$\frac{dO}{dt} = \alpha (1 - O) - \beta O$$

with  $\alpha = 10$ ,  $\beta = 1$  and  $O(0) = 0.5$

Choose an appropriate time step  $h$ !